**Thermodynamic analysis of the pathway for ethanol production from cellobiose in *Clostridium thermocellum***

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**Abstract**

**Background**

*Clostridium thermocellum* is an ideal candidate for consolidated bioprocessing by carrying out both cellulose degradation and fermentation. However, despite significant efforts the maximum ethanol titer achieved to date remains below industrially required targets. Several studies have analyzed the impact of increasing ethanol concertation on *C. thermocellum*’s membrane integrity, cofactor pool ratios, and altered enzyme regulation. In this study, we explore the extent to which thermodynamic equilibrium effects limit maximum ethanol titer.

**Results**

We use the max-min driving force (MDF) algorithm to identify the range of allowable metabolite concentrations that maintain a negative free energy of change for all reaction steps in the pathway from cellobiose to ethanol. In this study, we evaluate the thermodynamic consistency of the metabolomic dataset from the work by Tian et al 2017 [1] and then use it as constraint to demonstrate the thermodynamic favorability of pyruvate phosphate dikinase (PPDK) over malate shunt due to limiting CO2 concentrations and identify glyceraldehyde-3-phosphate dehydrogenase (GAPDH) as the key thermodynamic bottleneck under high external ethanol concentrations. The MDF approach was also deployed in a prospective mode to identify pathway modifications alleviating the thermodynamic bottleneck by analyzing EFMs with combinations of ten genetic interventions that are under consideration for improving ethanol titer in *C. thermocellum*.\.

**Conclusions**

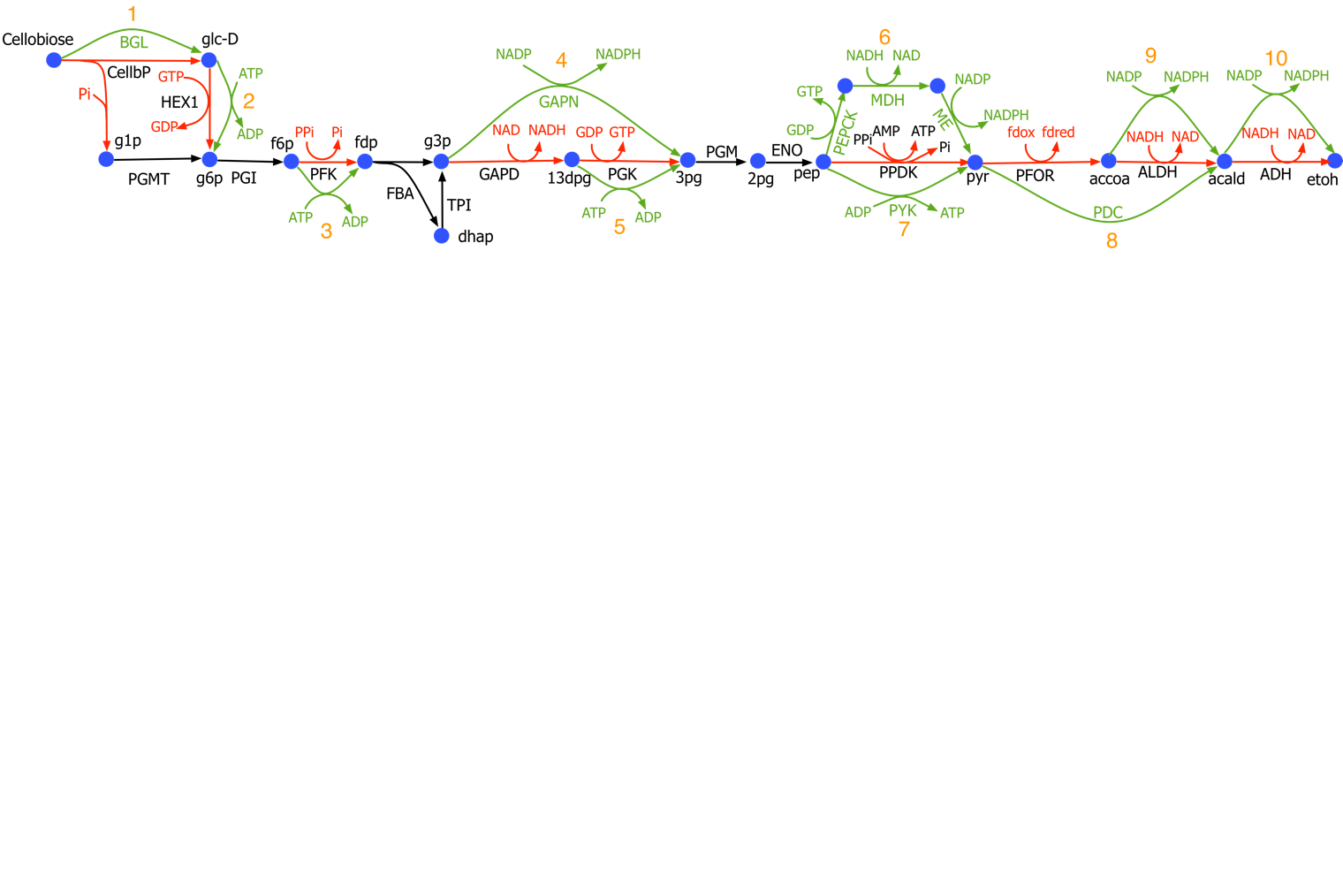
EFMs with NADPH dependent glyceraldehyde-3-phosphate dehydrogenase (GAPN) allow ethanol production at ethanol concentrations above 2M, however at the cost of eliminating net ATP production. and those with malate shunt had a significantly low MDF due to limiting CO2 concentrations under physiological conditions. For the set of interventions which allow net ATP production, ATP-linked phosphofructokinase (PFK-ATP) and NADPH linked alcohol dehydrogenase (ADH-NADPH) emerged as the best single genetic interventions maximize pathway MDF. This study shows that pathway thermodynamics can inform our understanding of metabolic phenotypes and propose testable solutions for restoring pathway thermodynamic feasibility.

**Background**

Biofuels have emerged as a major alternative to reduce fossil fuel emissions in the recent years [2-4]. One of the most industrially relevant biofuel production strategy relies on the microbial breakdown of cellulosic biomass, release of sugars and subsequent fermentation to alcohols [5]. *Clostridium thermocellum* is a promising candidate as it can both solubilize cellulose and produce ethanol. However, low ethanol titers [6] which have been linked to ethanol tolerance [1] have so far prevented its industrial adoption. Several studies have delved into problem of ethanol tolerance with focus on changes to *C. thermocellum*’s membrane integrity, varying cofactor pool ratios, and feedback regulations which affect enzyme kinetics [7-9]. An alternate approach to this problem as explored in this study involves identifying any limitations on the fundamental driving force of metabolism i.e. thermodynamics.

Thermodynamic constraints have already been integrated in computational modeling techniques such as flux balance calculations to determine possible reaction directionality and predict feasible metabolite concentration ranges [10]. Kinetic parameterization procedures often seek aid of thermodynamic constraints (i.e., negative free energy of change imperatives) to reduce the search space of feasible kinetic parameters and restrict reaction reversibility [11]. Thermodynamic analysis can be extended beyond single reaction to entire pathways. The thermodynamic feasibility of the operation of an entire pathway can be evaluated using the max-min driving force (MDF) formulation [12]. In MDF analysis, the concentrations of intracellular metabolites are allowed to vary to change the thermodynamic driving force of each reaction. By design, the solution identified by MDF is the one that maximizes the smallest driving force among all reaction steps (i.e., max-min formulation) akin to the optimization of the worst-case scenario [12]. Because metabolite concentrations are treated as optimization variables, the impact of metabolite pool accumulations and/or depletions on pathway feasibility can be directly assessed [12]. MDF has already been applied to study the causes of growth cessation in *C. thermocellum* under high substrate loading conditions, suggesting that fermentation is inhibited at the pyruvate to acetyl-coA conversion step due to increases in hydrogen (which inhibits the PFOR reaction via increased reduced ferredoxin levels) and increases in formate (which inhibits the PFL reaction) [9].

In this study, we use MDF to study the impact of increasing ethanol concertation on *C. thermocellum*’s metabolism. We restrict our analysis to glycolysis , the ethanol production pathway, (Figure 1) and its cofactor regeneration system. As input data, we use a set of previously-generated intracellular metabolite measurements from *C. thermocellum* cells grown with and without external ethanol addition [].We then explore ten plausible metabolic interventions (Summarized in Table 1) Several of these genetic interventions have already been explored in *C. thermocellum* with reported increase in ethanol titers [13-15], however the mechanism is not always fully known. In this study, we evaluate the extent to which those increases can be explained by changes in the thermodynamic driving force landscape. These genetic interventions, both individually and in combination,are evaluated by generating a set of unique flux solutions for cellobiose to ethanol conversion (i.e. elementary flux modes or EFMs) which contain one or more of the interventions. An EFM here represents a minimal set of reactions under steady state conditions with redox and energy balance while ensuring substrate (cellobiose) consumption and product (ethanol) formation [16]. We evaluate MDFs for 336 EFMs to give us a comprehensive view of the thermodynamic landscape, to allow us to prioritize future metabolic engineering work.

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**Figure 1:** Glycolysis with ethanol production pathway in C. thermocellum: Reactions and cofactors highlighted in black are present in all cases, those highlighted in red are replaced by those highlighted by green in the corresponding mutants, i.e. we replace 1) cellobiose hydrolase (CellbP) with betaglucosidase (BGL) 2) GTP/GDP with ATP/ADP as cofactors for gluckokinase 3) PPi/Pi with ATP/ADP as cofactors for PFK, 4) glyceraldehyde dehydrogenase (GAPD) and phosphoglycerate kinase (PGK) with glyceraldehyde-3-phosphate dehydrogenase (GAPN), 5) GTP/GDP with ATP/ADP as cofactors for phosphoglycerate kinase (PGK) , 6) PPDK with PEPCK and malate shunt, 7) PPDK with PYK, 8) pyruvate formate oxidoreductase (PFOR) and aldehyde dehydrogenase (ALDH) with pyruvate decarboxylase (PDC), 9) NADH/NAD with NADPH/NADP as cofactors for aldehyde dehydrogenase, and 10) NADH/NAD with NADPH/NADP as cofactors for alcohol dehydrogenase

**Table 1.** Reaction in Wild-type *C. thermocellum* and the corresponding replacements

|  |  |  |  |
| --- | --- | --- | --- |
| Wild-type | | Replaced with | |
| Reaction name | Equation | Reaction name | Equation |
| CBP | pi + cellb <=> glc-D + g1p | BGL | cellb + h2o <=>2 glc-D |
| GLK-GTP | glc-D + gtp <=> g6p + gdp | GLK-ATP | glc-D + atp <=> g6p + adp |
| PFK-PPi | ppi + f6p <=> pi + fdp + h | PFK-ATP | atp + f6p <=> adp + fdp |
| GAPDH | pi + nad + g3p <=> nadh + 13dpg | GAPN | g3p + nadp + h2o <=> 3pg + nadph |
| PGK-GTP | gdp + 13dpg <=> gtp + 3pg |
| PGK-GTP | gdp + 13dpg <=> gtp + 3pg | PGK-ATP | adp + 13dpg <=> atp + 3pg |
| PPDK | amp + ppi + pep <=> atp + pi + pyr + h | PYK | adp + pep <=> pyr + atp |
| PPDK | amp + ppi + pep <=> atp + pi + pyr + h | PEPCK | gdp + co2 + pep <=> gtp + oaa |
| MDH | nadh + oaa <=> nad + mal-L |
| ME | nadp + mal-L <=> nadph + co2 + pyr |
| PFOR | coa + 2 fdxox + pyr <=> h + accoa + 2 fdxrd + co2 | PDC | pyr <=> acald + co2 |
| ALDH-NADH | nadh + accoa <=> nad + coa + acald |
| ALDH-NADH | nadh + accoa <=> nad + coa + acald | ALDH-NADPH | nadph + accoa <=> nadp + coa + acald |
| ADH-NADH | acald + nadh <=> etoh + nad | ADH-NADPH | acald + nadph <=> etoh + nadp |

# Results and discussion

## Determining the cause of ethanol inhibition

In a previous study, the addition of ethanol was shown to inhibit *C. thermocellum* metabolism at the GAPDH reaction [1]. Although Tian et al. suggested that the inhibition was due to regulatory behavior, an alternative hypothesis is that the inhibition was due to one or more reactions reaching thermodynamic equilibrium. To test this hypothesis, we re-analyzed the data from Tian et al. using the MDF thermodynamic analysis framework developed by Noor et al. ().

This dataset represents intracellular metabolites collected from wild type (WT) *C. thermocellum* growing with and without (control) the presence of added ethanol for two replicates at three different timepoints. The ethanol concentration is steadily increased to reach a maximum concentration (i.e. at the final timepoint) of 40 g/L, the point at which growth is inhibited. The depletion of extracellular cellobiose and increase in fermentation products, such as lactate, acetate, ethanol (Supplementary table FFF), shows that cellobiose was converted to ethanol during all three timepoints for the no-ethanol control and for the first two timepoints for the ethanol added samples implying thermodynamic feasibility of ethanol production for those timepoints. However, for the case of samples with added ethanol, we observe reduced consumption of cellobiose and the lack of increase in ethanol pool for the final timepoint indicating a lack of flux from cellobiose to ethanol (which could be due to a regulatory event or thermodynamic equilibrium). If the pathway inhibition is due to a regulatory event, we would expect the ∆Gr for the GAPDH reaction to have a large negative value. If the pathway inhibition is due to thermodynamic equilibrium, we would expect the ∆Gr to have a value close to zero. In theory, we could determine the ∆Gr of the GAPDH reaction by knowing the absolute concentrations of the reactants (NAD+, Pi and gap) and the products (NADH and 3pg), however the Tian et al. dataset only has data for some of those metabolites. By building a thermodynamic model of the entire pathway, we can use data from other measured metabolites to further constrain the system. In addition, we need to identify reasonable concentration ranges for non-measured metabolites.

Thus, the goal of our initial analysis was to determine a set of model parameters that are thermodynamically feasible (i.e. positive MDF value) for all 3 timepoints of the no-ethanol control, and the first 2 timepoints of the ethanol-addition condition. For each measured metabolite, we include an uncertainty factor of 1.5 (see materials and methods for description) to account for quantification error. Starting with the default metabolite concentration bounds proposed by Noor et al., we relaxed several constraints (Table iii) to allow the model MDF predictions to match our known feasibility conditions.

**Table iii. Relaxation of metabolite bounds proposed by Noor et al.**

|  |  |  |  |
| --- | --- | --- | --- |
| **Constraint** | **Noor et al.** | **This work** | **Note** |
| Default bounds | 1 µM to 10 mM | 1 µM to 20 mM | Measured concentrations for some metabolites are > 10 mM |
| ATP/ADP ratio | 10 | 10-200 |  |
| GTP/GDP ratio | 10 | 10-200 |  |
| NAD/NADH ratio | 10 | 0.01-100 | This ratio is constrained by thermodynamics of GAPDH, ALDH and ADH reactions |
| NADP/NADPH | 0.1 | 0.01-100 |  |
| Fd(ox)/Fd(red) | 1 | 0.01-100 |  |
| PPi | 1 mM | 1 µM to 20 mM |  |
| CoA | 1 mM | 1 µM to 20 mM |  |

Next, we evaluated measured metabolites one at a time. The results indicate that DHAP and FDP measurements are thermodynamically inconsistent resulting in negative MDF for all timepoints (Supplementary table GGG). DHAP is difficult to measure by mass spectrometry because it has the same mass to charge (M/z) ratio as G3P, and the two metabolites are interconverted by the TPI reaction, potentially allowing for rapid equilibration during quenching. Because of these uncertainties, we suspect the measurement error for DHAP may be higher than 1.5. FDP is readily measured by the LC/MS system we are using [], so its lack of thermodynamic feasibility is puzzling. In the no-added-ethanol samples, the FDP levels are around 10 µM, however values would need to be 5-fold higher (at least 50 µM) to ensure thermodynamic feasibility. Although resolving this discrepancy is not the main point of this work, we recognize that this is an important task for future metabolomic studies in *C. thermocellum*. Measured values for DHAP and FDP were ignored for subsequent analyses.

Finally, we performed our MDF analysis using all of the remaining measured metabolite values as constraints (ethanol, 3pg, accoa, f6p, g6p, g1p, glc-d, pep, pyr and mal-l) (note that cofactors were not included, see materials and methods for detailed explanation). The complete model output (metabolite concentrations, ∆Gr and MDF values for each condition and timepoint is presented in supplemental figure JJJ). For all of the “feasible” conditions, the reaction shadow price is concentrated in the PGI reaction. This is because this is the only reaction whose ∆Grvalue is completely determined by measured metabolite values (i.e. g6p and f6p). For the “infeasible” condition (40 g/L added ethanol), the reaction shadow price is concentrated in the CBP reaction and the metabolite shadow prices are concentrated in cellobiose and inorganic phosphate (neither of which are measured). What this means is that at 40 g/L added ethanol, the maximum ethanol titer is determined by the cellobiose concentration, and thus we cannot exclude the possibility that ethanol production stops due to the attainment of thermodynamic equilibrium across a large portion of the reactions in the cellobiose to ethanol pathway (including GAPDH). It must be emphasized that this result is partly due to the lack of model constraints. Increasing the number of measured metabolites (particularly cofactors, which participate in many reactions), determining the problems with DHAP and FDP quantification, and improving the accuracy of cofactor quantification (which would reduce the error factor below the current value of 1.5) will further improve our understanding of the thermodynamic landscape. Nevertheless, we can identify feasible ranges for several cofactors.

**Cofactor ratios**

NAD/NADH ratio -

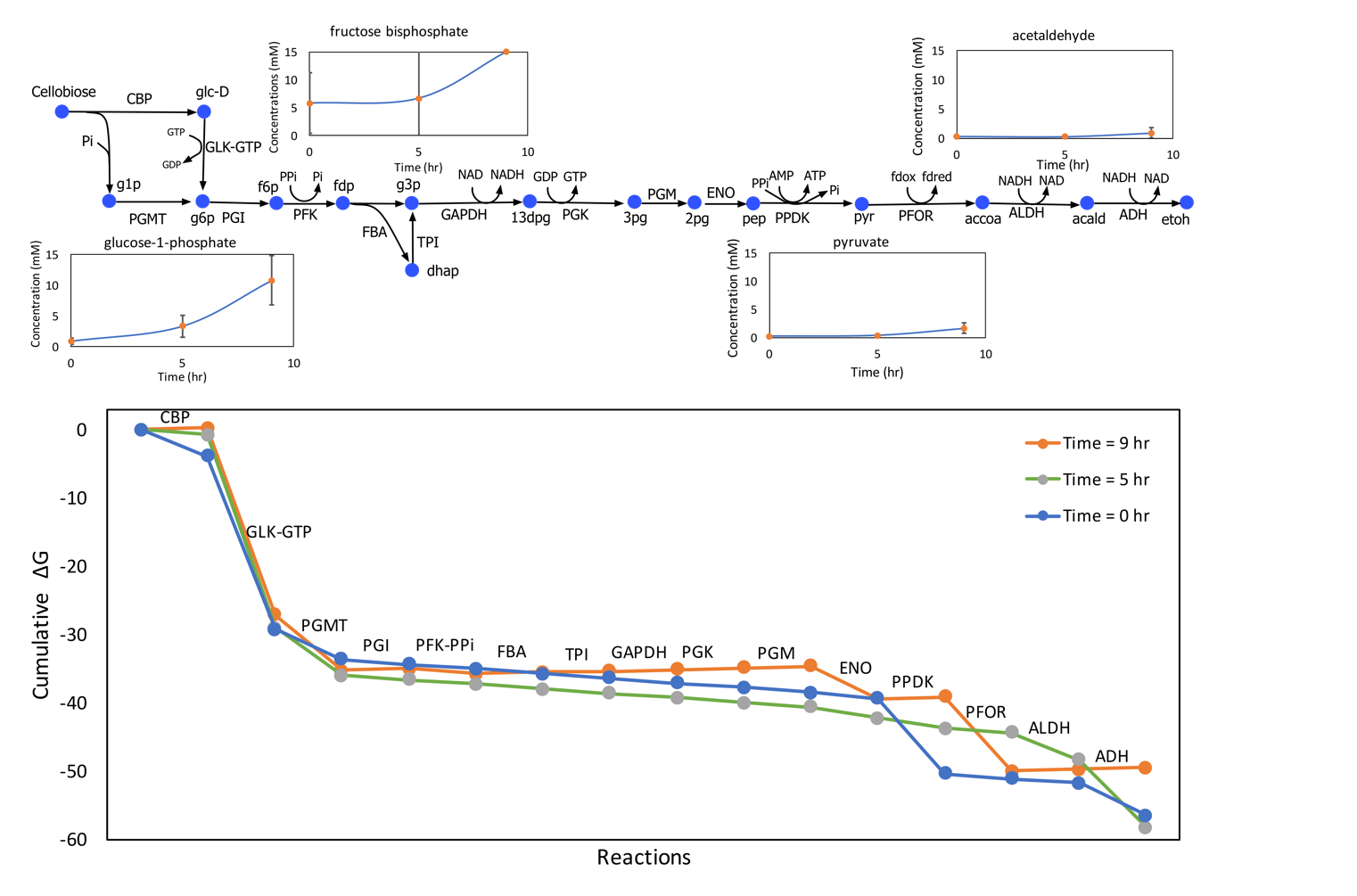
ATP/ADP ratio – not constrained. This ratio is thought to be high to support biosynthetic reactions, which are not accounted for in this model.

GTP/GDP ratio – same as ATP/ADP ratio.

PPi/Pi ratio – if this ratio is too low, the PPi-PFK reaction doesn’t work.

**Table 2.** Pathway MDF of wild-type *C. thermocellum* for all samples and time points with and without added ethanol. Negative MDF values indicate thermodynamic infeasibility at the final timepoints of ethanol addition

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Time point (hr) | Externally added ethanol | | No added ethanol | |
| Sample 1 | Sample 2 | Sample 3 | Sample 4 |
| 0 | 1.08 | 0.69 | 0.78 | 0.50 |
| 5 | 0.03 | 0.66 | 1.17 | 0.85 |
| 9 | -0.22 | -0.23 | 1.05 | 0.70 |



**Figure 2:** Glycolysis with ethanol production pathway in wild-type C. thermocellum with PPDK showing the variation in metabolite concentrations of key metabolites across time points. The ethanol concentration increases over time due to the addition of ethanol. The accumultation of sugar phosphates (e.g. G1P) in upper glycolysis is correlated with increased ethanol titer.

## Analysis of possible metabolic engineering interventions in *C. thermocellum*

[think of good transition that links thermodynamic equilibrium argument from previous section EFM analysis in this section]

MDF analysis of wild-type *C. thermocellum* in the previous section showed that cofactor pools (especially NADH) plays a major role in driving the thermodynamic feasibility of ethanol production in *C. thermocellum*. Thus, we can infer that perturbations in cofactor pools can be used to our advantage to increase ethanol production by ensuring a larger MDF for the pathway. To this end, we have systematically evaluated pathway modification strategies by genetic interventions which modify the cofactor association of a single or multiple reactions in *C. thermocellum* including: beta-glucosidase (BGL), ATP-linked phosphofructokinase (ATP-PFK), NADP-dependent glyceraldehyde 3-phosphate dehydrogenase (GAPN), pyruvate decarboxylase (PDC), NADPH-linked ALDH (ALDH-NADPH), and NADPH-linked ADH (ADH-NADPH) (see Figure 1 and Table 1). A genetic intervention often implies multiple changes. For example, changing the cofactor preference of the ADH reaction from NADH to NADPH implies a second change somewhere in the pathway to provide a source of NADPH. To ensure that all of these secondary changes were accounted for, we generate elementary flux modes (EFMs) for all combinations of reactions that allow conversion of one molecule of cellobiose into four molecules of ethanol while allowing for only unique biotransformations (i.e. prevent reactions which differ only in cofactors to appear in the same EFM). For each of the resulting 336 EFMs (Supplementary Table XX), we calculate the ATP generated and the pathway MDF. We quantify the difference between our wild-type EFM and any mutant EFM by calculating their Hamming Distance which measures the number of reaction changes in a mutant compared to our wild-type EFM. A high hamming distance indicates that significant strain design effort would be invested in generating the mutant strain. All the 336 EFMs are analyzed at 1M ethanol concentration while allowing other metabolite concentrations to vary within physiological range (1 μM - 0.02 mM), the energy cofactors (ATP/ADP, GTP/GDP, and PPi/Pi) were allowed to assume a ratio greater than 10:1 and the redox cofactors (NADH/NAD+, NADPH/NADP+, Fd(red)/Fd(ox)) ratios were allowed to vary between 1:100 and 100:1. Figure 3 summarizes the MDF values, ATP generation, and hamming distance of all the EFMs.

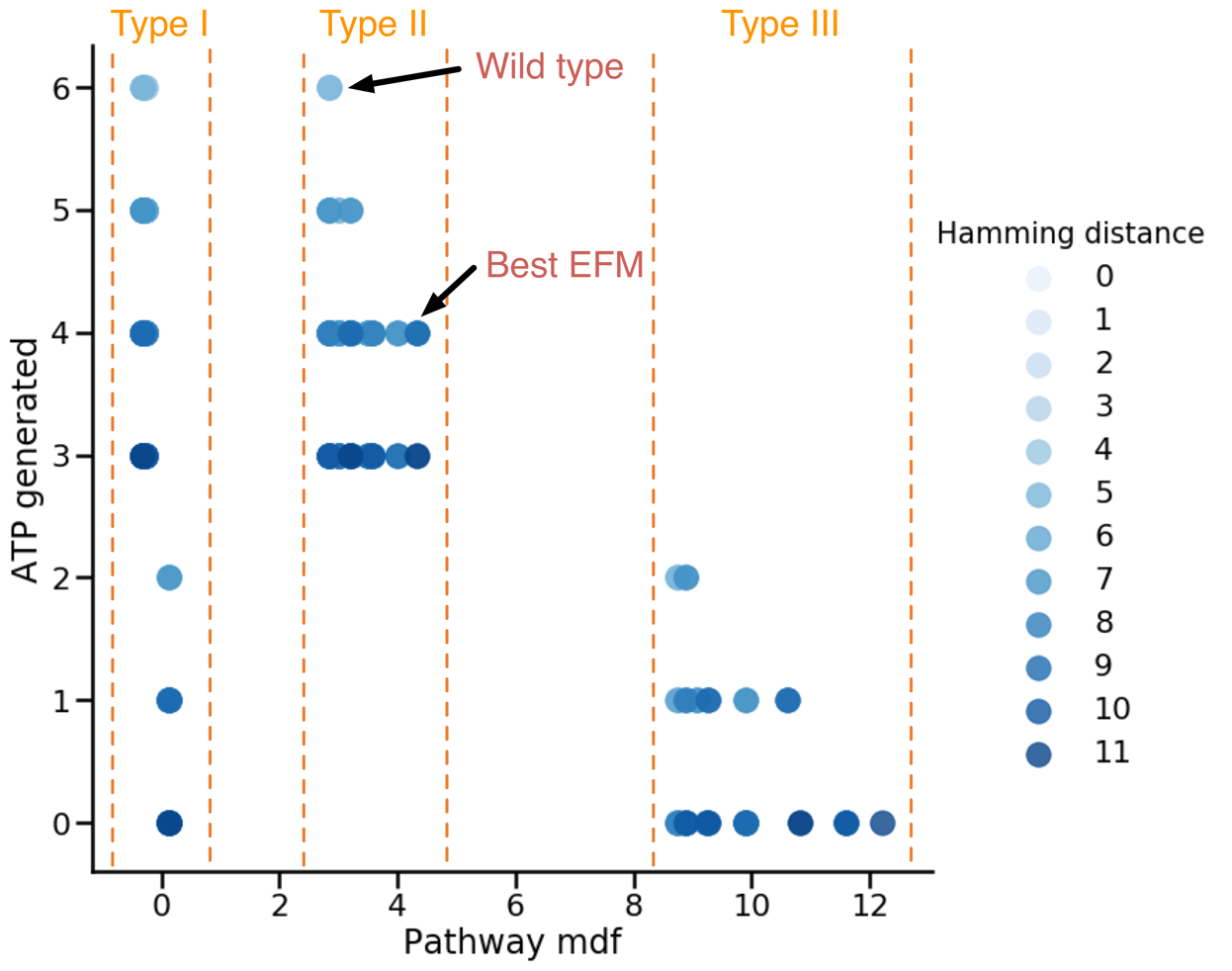
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Figure 3: MDF at 1M ethanol concentration, ATP generated, and Hamming distance of all 336 EFMs. The EFMs are generated using all possible combinations of cofactor modifications as list in Table 1. The shade of blue represents the value of hamming distance of any given EFM calculated from the wild-type EFM. The EFMs are clustered in three different regions based on their MDF values.

There are three distinct clusters of EFMs which can be seen in Figure 3 based on pathway MDFs, i.e. type I EFMs with low MDF (<1), type II EFMs with intermediate MDF (3-5) but high ATP generation (>2), and type III EFMs with high MDF (>8) but low ATP generation (<3). It is interesting to note that all the type I EFMs possessed the malate shunt to generate pyruvate which led to thermodynamic infeasibility except those EFMs which additionally contain the GAPN reaction. The physiologically allowable CO2 concertation restricted the PEPCK reaction lowering the MDF for all these EFMs close to zero indicating poor thermodynamic feasibility similar to the case with WT *C. thermocellum* as discussed in the previous section. All the type III EFMs possessed the GAPN reaction which replaces GAPDH and PGK reactions from wild-type *C. thermocellum*. Studies have shown that such a genetic intervention can improve ethanol yield in yeast and lysine yield in *Corynebacterium glutamicum* [22, 23]. However, introduction of GAPN also reduces the ATP generated by the pathway which is necessary for cell growth. The EFM with highest MDF completely removed NADH association with the ethanol production by using PDC and NADPH linked ADH. PDC replaces PFOR and ALDH-NADH and hence rids the pathway of redox regeneration systems which entail generation of NADPH or NADH from reduced ferredoxin. The driving force advantage gained by GAPN and PDC can be also understood by the fact that they reduce either the energy generation (GTP in case of PGK) or the redox generation (reduced ferredoxin by PFOR) of the native pathway which negatively impacts pathway MDF. It is noteworthy that studies have shown a higher NADPH/NADP ratio compared to NADH/NAD ratio in *C. thermocellum* and thus NADPH linked reactions would tend to have a stronger driving force in engineered strains [24].

The most useful genetic interventions are part of the type II EFMs which retain ATP generation along with intermittent pathway MDF as shown in Figure 3. All the high MDF type II EFMs possess the ATP linked PFK reaction and the NADPH linked ADH reaction. The best EFM has a minimum hamming distance of 2 indicating PFK-ATP and ADH-NADPH as the only changes from native pathway. The ATP-PFK provides a stronger driving force at the cost of ATP generation (PPi is equivalent to 0.5 ATP [25]) while the NADPH linked ADH reaction decouples the impact of rise in ethanol concentrations from NADH associated native reactions. Studies have also shown that the cofactor specificity of alcohol dehydrogenase changes to NADPH from NADH in high ethanol yielding strains of *C. thermocellum* [26]. We can thus conclude that ATP-PFK and NADPH-ADH are the most beneficial genetic intervention which can help us achieve high ethanol titers while sustaining cell growth.

## PPDK vs malate shunt

*C. thermocellum* does not possess the conventional pyruvate kinase (PYK) enzyme, instead it uses pyruvate phosphate dikinase (PPDK) and phosphoenolpyruvate carboxykinase (PEPCK) with the malate shunt as two alternate pathways to generate pyruvate from phosphoenolpyruvate (see Figure 1). The metabolic flux allocation between these two pathways is essential to characterize wild-type metabolism and predict the impact of genetic perturbations on metabolism using kinetic models. Several studies have demonstrated that the flux ratio between PPDK and malate shunt is tilted heavily towards PPDK [8, 13] suggesting the usefulness of malate shunt only for its transhydrogenase activity for anabolic processes [20]. In this study, we evaluate the driving force for the two pathways using wild-type measurements under varying ethanol concentrations. The results (shown in Figure 2) clearly indicate that the thermodynamic driving force is consistently positive for PPDK for ethanol concentrations up to 0.8M in contrast to the malate shunt which remains thermodynamically infeasible for the entire range of ethanol concentrations (0-1M). This is because the malate shunt requires a high intracellular CO2 concentration (>0.1M) to ensure thermodynamic feasibility, but the CO2 concentration bounds are restricted (<0.01mM) under on standard experimental conditions [12]. This alludes to a localized increase of CO2 concentration in WT *C. thermocellum* for PEPCK to be feasible and also indicates close proximity of PEPCK to CO2 generating reactions such as pyruvate: ferredoxin oxidoreductase (PFOR), malate dehydrogenase (MDH) which should be probed experimentally. We also observe accumulation of upper glycolysis metabolites especially sugar phosphates (also observed in ethanol stress studies [1, 21]) as a consequence of GAPDH being a thermodynamic bottleneck due to an increase in NADH pool under ethanol stress (Figure 2).

**Conclusions**

This study analyzes wild-type *C. thermocellum* glycolysis using experimentally measured concertation datasets to understand the impact of increasing ethanol concertation. GAPDH was revealed as the major thermodynamic bottleneck at high ethanol concentrations which renders ethanol production infeasible. EFMs of plausible genetic interventions showed the infeasibility of malate shunt under physiological CO2 concentrations and the benefits of having the GAPN reaction which generates high MDF by resolving the GAPDH bottleneck but with reduced ATP generation at high ethanol concentrations. The best genetic intervention which retains ATP generation with a high driving force is the combination of ATP linked PFK and NADPH linked ADH reaction which has also been shown to occur in high ethanol yielding *C. thermocellum* strains [26].

Conventional studies have explored high yield target phenotypes by manipulating networks to improve desired fluxes. This study shows that a fundamental analysis of pathway thermodynamics can augment our arguments for a given genetic perturbation by providing a mechanistic detail of the mutant phenotype and help us weed out thermodynamically infeasible designs in our search for overproducing strains. However, we should be careful about the simplified assumptions which were essential for this study but would play vital role in real world scenarios. Cellulose degradation has not been considered in this study, but there are several challenges associated with this step including enzyme kinetics, cellulosome availability, and most of all recalcitrance. Enzyme kinetics are also essential to accurately estimate the impact of changes in metabolite and regulator pools on reaction fluxes through allosteric and feedback regulations. In this study, we also assumed the intracellular concentration of ethanol to be the same as the extracellular concentration, since the cytoplasmic membrane is not thought to be a barrier to ethanol diffusion [27], however, this assumption may not hold true for other potential biofuels such as butanol and would require intracellular measurements for accurate quantification. Despite all these simplifications, our results show that thermodynamic pathway analysis remains a vital tool to prune potential strain design strategies.

# Materials and methods

## Metabolite quantification (Texts from Dan)

A single 200 ml culture was grown to an OD600 of 0.1, the culture was split in half. Starting at 2 hours (post-split), ethanol was added to one culture at a rate of about 9 g/L/h. No ethanol was added to the other culture. At three timepoints (T=2.0h, 3.8h and 5.9h), each culture was sampled twice for intracellular metabolites using previously described protocols [1, 24, 28], and once for extracellular metabolites (supplemental table BBB). The raw data was re-processed with El-Maven 0.5.0 and quantified using external standards (supplemental figure AAA). Since the response was not linear over the full range of the standards (0.1 µM to 100 µM), quantification was performed by piecewise linear interpolation (supplemental figure DDD). Based on four measurements of our standard curves, we used an uncertainty factor of 1.5 during our simulations. One significant change that we observed from the absolute quantification is that the energy charge (ATP or GTP, calculated using Equation (1)) values which had previously been calculated to be around 0.9, were actually much lower, in the range of 0 to 0.15, which is much lower than expected [28], and thus datasets with 0 energy charge were excluded from analysis.

(1)

In addition, NADH was not observed for the no ethanol control cultures. Since NADH and NAD+ form a cofactor pair, both metabolites were excluded from analysis. Since metabolites are typically diluted during the quenching and extraction process, to determine the *intracellular* concentration of metabolites, the measured concentration was adjusted using Equation (2).

(2)

The intracellular volume is assumed to be 3.9 µL for 1 ml of a culture at an OD600 density of 1 [29]. The concentration factor was calculated separately for each extraction, based on the measured OD600 and sample volume. The target cell density (OD600 x ml) was 2.0, but varied from 1.6 to 2.1 (supplemental table CCC). For ethanol, the intracellular concentration was assumed to be the same as the extracellular concentration, since the cytoplasmic membrane is not thought to be a barrier to ethanol diffusion [27].

## Assessing the thermodynamic feasibility of a pathway

The thermodynamic feasibility of a given pathway is assessed using the max-min driving force (MDF) formulation [12]. The MDF formulation identifies a set of metabolite concentrations that ensure the lowest free energy changes for all the reactions in a pathway. The MDF problem minimizes the maximum of a pathway by optimizing over the concentrations of all metabolites in the pathway. The formulation is given by:

|  |  |  |
| --- | --- | --- |
|  | (-MDF) | (3) |
|  |  | (4) |
|  |  | (5) |
|  |  |  |

where *I*is the set of all metabolites and *J*is the set of all reactions in a given pathway, is the concentration of metabolite , is the gas constant, is the temperature and the matrix refers to the stoichiometric matrix of the pathway with . Constraint (4) relates the Gibbs free energy of reaction () with the standard Gibbs free energy of reaction () and the mass action ratio. The pathway with a positive objective function (i.e. negative MDF) indicates that it is thermodynamically infeasible within the given physiological concentration (and ratio) ranges. The MDF problem is solved using Gurobi Optimizer v6.5.1 solver and Python script modified from the Equilibrator-API Python package [30].

For performing the max-min driving-force (MDF) analysis, maximum and minimum metabolite concentration have to be established for non-measured metabolites. Noor et al [12] proposed a range of 1 µM to 10 mM, based largely on the work of Bennet et al [18]. Based on our measurements, we decided to keep the lower default concentration at 1 µM, but raise the upper default concentration from 10 mM to 20 mM. Although Noor et al fixed the ratios of several cofactor pairs, we have relaxed these constraints, since the values have not been experimentally determined for *C. thermocellum*. Several cofactors were excluded from analysis due to measurement problems (see Metabolite quantification section). To avoid biasing the results by including only a few measured cofactors, we opted to ignore measured values for all cofactors (ATP, ADP, AMP, GTP, GDP, NAD+. NADH, NADP+ and NADPH). Concentration ranges for energy cofactors (ATP/ADP, ATP/AMP, GTP/GDP) were set to allow ratios greater than 10:1 and redox cofactors (NADH/NAD+, NADPH/NADP+, Fd(red)/Fd(ox)) to vary between 1:100 and 100:1. Default bounds for each metabolite is described in Supplementary table EEE.

## EFM evaluation

We implement the algorithm to generate k-shortest EFM of a network to systematically evaluate all the EFMs associated with our network [31]. The formulation is given by:

|  |  |  |
| --- | --- | --- |
|  |  | (6) |
|  |  | (7) |
|  |  | (8) |
|  |  | (9) |
|  |  | (10) |
|  |  | (11) |
|  |  | (12) |

where *I* is the set of all metabolites and *J* is the set of all reactions, is the set of exchange reactions, the rate of reactions is represented by and represents the stoichiometric coefficient of the metabolite *i* in reaction *j* reactions in a given pathway, *K* represents the set of previously found solutions and indicates the optimal value of the binary variable in the *kth* solution . The binary variable (equation 10) assumes a value of 1 if the reaction *j* is included in the EFM and 0 otherwise. The smallest EFM is found by minimizing the sum of all these binary variables as shown by constraint 6. Steady state of the network is ensured by enforcing no net accumulation or consumption of metabolites as shown by constraint 7. The network is decomposed to ensure that all reaction fluxes are positive (constraint 11) and the binary variable was linked to reaction rates by constraint 8. The binary variables corresponding to exchange reactions () are fixed by constraint 9 to ensure that all EFMs consume the substrate (cellobiose) and generate the product (ethanol). We finally use integer cuts (constraint 12) to generate all possible EFMs associated with the network. The hamming distance between the wild-type and any given EFM is calculated by calculating the number of wild-type reaction which are absent in the mutant EFM. The ATP generated by any given EFM is calculated by adding the fluxes of the two ATP hydrolysis reaction in the models (Table 3).

Table 3: List of all possible reactions in any given EFM.

|  |  |
| --- | --- |
| **Reaction Name** | **Equation** |
| ATPase1 | h2o + atp <=> adp + pi |
| CBP | pi + cellb <=> glc-D + g1p |
| BGL | cellb + h2o <=>2 glc-D |
| GLK-GTP | glc-D + gtp <=> g6p + gdp |
| GLK-ATP | glc-D + atp <=> g6p + adp |
| PGMT | g1p <=> g6p |
| PGI | g6p <=> f6p |
| PFK-PPi | ppi + f6p <=> pi + fdp + h |
| PFK-ATP | atp + f6p <=> adp + fdp |
| FBA | fdp <=> dhap + g3p |
| TPI | dhap <=> g3p |
| GAPDH | pi + nad + g3p <=> nadh + 13dpg |
| GAPN | g3p + nadp + h2o <=> 3pg + nadph |
| PGK-ATP | adp + 13dpg <=> atp + 3pg |
| PGK-GTP | gdp + 13dpg <=> gtp + 3pg |
| PGM | 3pg <=> 2pg |
| ENO | 2pg <=> pep + h2o |
| PYK | adp + pep <=> pyr + atp |
| PPDK | amp + ppi + pep <=> atp + pi + pyr + h |
| PEPCK | gdp + co2 + pep <=> gtp + oaa |
| MDH | nadh + oaa <=> nad + mal-L |
| ME | nadp + mal-L <=> nadph + co2 + pyr |
| PFOR | coa + 2 fdxox + pyr <=> h + accoa + 2 fdxrd + co2 |
| RNF\_ATPase | 2 fdxrd + 1 nad + 0.25 adp + 0.25 pi <=> 2 fdxox + 1 nadh + 0.25 atp + 0.25 h2o |
| RNF\_PPiase | 2 fdxrd + 1 nad + 1 h + 1 pi <=> 2 fdxox + 1 nadh + 0.5 ppi + 0.5 h2o |
| NFN | 2 fdxrd + nadh + 2 nadp = 2 fdxox + nad + 2 nadph |
| PDC | pyr <=> acald + co2 |
| ALDH-NADH | nadh + accoa <=> nad + coa + acald |
| ALDH-NADPH | nadph + accoa <=> nadp + coa + acald |
| ADH-NADH | acald + nadh <=> etoh + nad |
| ADH-NADPH | acald + nadph <=> etoh + nadp |
| Bifur\_Hyd | 2 fdxrd + nadh + 2 h = nad + 2 h2 + 2 fdxox |
| NDK | gtp + adp <=> atp + gdp |
| Gly-cyc | atp + pi <=> adp + ppi |
| NADPH-FNOR | 2 fdxrd + nadp <=> 2 fdxox + nadph |
| NADH-FNOR | 2 fdxrd + nad <=> 2 fdxox + nadh |
| ATPase2 | atp + h2o <=> amp + ppi |

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